Synthesis, antifungal and nematocidal activities of thioureines with an aminoester sequence

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Summary — Twenty-three arylthioureines bearing β -alanine or γ -aminobutyric alkyl ester chains were synthesized for *in vitro* screening toward 44 strains of fungi and 2 genera of nematodes. The nitro derivatives were the most potent compounds against *Aspergillus* and *Candida* strains. Ester chains increase activity against the filarial worm *Molinema dessetae*. Twelve compounds have $EC_{50} < 40 \,\mu\text{g/ml}$. However, the anthelmintic potency is weak compared with tetramisole.

antifungal activity / anthelmintic / thioureine

Introduction

Since the discovery of anthelmintic tetramisole 1 (fig 1), a number of analogues have been synthesized, particularly 6-(3-aminophenyl) derivatives. The corresponding thioureas have been reported to possess broad spectrum activity against nematodes, trematodes and cestodes [1]. Many authors have reported anthelmintic activity for molecules bearing the thiourea moiety [2–4]. Walchshofer et al [5] have studied structural parameters for thiazolyl ureas and thioureas and noted the influence of lipophilicity on anthelmintic activity. In a factorial analysis of the correspondence between chemical structure and antiparasitic activity, Doré et al [6] revealed the potential of thioureas as antiparasitic agents. Numerous sulfurcontaining molecules with divalent sulfur attached to carbon atoms have in vitro fungistatic action, eg, dithiocarbamates and thiurams [7]. Thus, thioureines are analogues of antifungal structures and were tested against fungi.

On basis of these results, we previously studied aminothiazolines 2 and aminothiazines 3 [8], which are open analogues of anthelmintic tetramisole 1 (fig 1), and then thioureines 4 and 5 bearing a β -alanine or γ -aminobutyric moiety [9]. Such structures are GABA-like compounds (GABA = γ -aminobutyric acid) and potential anthelmintic agents,

because GABA is an inhibitory transmitter for the neuromuscular system of nematodes [10]. GABA itself is not very potent and the moderate anthelmintic and antifungal activities of compounds 4 and 5 were assigned to their weak lipophilicity. In a continuation of this work, we report here the synthesis and activities of some more lipophilic esters of thioureines with the same β -alanine or γ -amino butyric sequences.

Chemistry

Compounds 9 were obtained in four steps from aromatic amines through condensation of corresponding isothiocyanates 8 with the appropriate aminoesters 6 or 7, following scheme 1. Isothiocyanates 8 were prepared as previously described by Hodgkins et al [11]; aminoesters of β -alanine or GABA were obtained in the usual manner. Physicochemical and NMR data for compounds 9a-w are given in tables I and II

Results

Antifungal activities

All compounds were screened *in vitro* against an array of clinical isolates of 35 strains of *Candida* and one reference strain *C albicans* AFNOR ATCC 2094, and

Fig 1. Structures of compounds 1–7.

Scheme 1.

eight strains of opportunistic fungi (Aspergillus fumigatus, A flavus, A niger and Scopulariopsis brevicaulis). The results are reported in tables III and IV. When the minimum inhibitory concentrations (MIC) are close for different strains of the same species, we report a median value.

Anthelmintic activities

The same compounds were tested against infective larvae of an intestinal parasite of rats, Nippostrongylus brasiliensis, and against infective larvae of a filaria, Molinema dessetae. These two tests were chosen because they tend to detect in vitro activities that are generally confirmed in vivo [6–8]. The results $(EC_{50} \text{ in } \mu\text{g/ml})$ are reported in table V.

Discussion

All the antifungal activities were moderate, except for 9d (4 µg/ml < MIC < 8 µg/ml for all strains of C albicans and 2 µg/ml < MIC < 8 µg/ml for all strains of Aspergillus). The two nitro compounds 9s and 9t were equipotent (4 µg/ml < MIC < 16 µg/ml for all strains of Candida in Casitone (CAS) medium and for all strains of Aspergillus, in both yeast nitrogen-based glucose (YNBG) and CAS medium) and their MIC values were less than econazole for all strains of Candida. These methyl and isopropyl esters indicate that the nature of the ester group is not determinant for activity. Moreover, three esters of the same molecule 9a—c had differing antifungal activity (9a < 9b < 9c); 9d is very active whereas the corresponding ethyl

Table I. Physicochemical data for compounds **9a-w**.

Ar—NH—C—NH—
$$(CH_2)$$
 $\frac{1}{n}$ COOR

Compound	Ar	n	R	Formula	MW	Mp (°C)	Yield (%)
9a	Phenyl	2	Methyl	$C_{11}H_{14}N_2O_2S$	238.2	72–73	80
9b	Phenyl	2	Ethyl	$C_{12}H_{16}N_2O_2S$	252.2	108-109	40
9c	Phenyl	2	<i>i</i> -Propyl	$C_{13}H_{18}N_2O_2S$	266.2	111-113	72
9d	p-Anisyl	2	Methyl	$C_{12}H_{16}N_2O_3S$	268.2	135	78
9e	p-Anisyl	2	Ethyl	$C_{13}H_{18}N_2O_3S$	282.3	138–139	46
9 f	p-Anisyl	2	<i>i</i> -Propyl	$C_{14}H_{20}N_2O_3S$	296.3	95-97	81
9g	p-Nitrophenyl	2	Methyl	$C_{11}H_{13}N_3O_4S$	283.3	153–154	38
9h	3,4-Dimethoxyphenyl	2	Methyl	$C_{13}H_{18}N_2O_4S$	298.3	123-124	51
9i	3,4-Dimethoxyphenyl	2	Ethyl	$C_{14}H_{20}N_2O_4S$	312.3	138–140	43
9j	3,4-Dimethoxyphenyl	2	i-Propyl	$C_{15}H_{22}N_2O_4S$	326.3	126	50
9k	2-Benzodioxanyl	2	Methyl	$C_{13}H_{16}N_2O_4S$	296.2	130	79
91	2-Benzodioxanyl	2	i-Propyl	$C_{15}H_{20}N_2O_4S$	324.3	114	40
9m	Phenyl	3	Methyl	$C_{12}H_{16}N_2O_2S$	252.2	131–132	59
9n	Phenyl	3	Ethyl	$C_{13}H_{18}N_2O_2S$	266.2	98-101	40
90	Phenyl	3	i-Propyl	$C_{14}H_{20}N_2O_2S$	280.2	107–109	75
9p	p-Anisyl	3	Methyl	$C_{13}H_{18}N_2O_3S$	282.3	112–114	58
9q	p-Anisyl	3	Ethyl	$C_{14}H_{20}N_2O_3S$	296.3	96	78
9r	p-Anisyl	3	<i>i</i> -Propyl	$C_{15}H_{22}N_2O_3S$	310.3	105-108	57
9s	p-Nitrophenyl	3	Methyl	$C_{12}H_{15}N_3O_4S$	297.3	155–157	53
9t	p-Nitrophenyl	3	i-Propyl	$C_{14}H_{19}N_3O_4S$	325.3	160-162	61
9u	3,4-Dimethoxyphenyl	3	i-Propyl	$C_{16}H_{24}N_2O_4S$	340.3	112	86
9v	2-Benzodioxanyl	3	Methyl	$C_{14}H_{18}N_2O_4S$	310.3	122	45
9w	2-Benzodioxanyl	3	i-Propyl	$C_{16}H_{22}N_2O_4S$	338.3	120-121	40

and isopropyl esters 9e and 9f have similar activities; and esters 9h–j of the same molecule have comparable levels of activity. It is thus impossible to correlate activity with the nature of the ester group. The influence of the nature of the aminoester group did not prove any superiority of β -alanine over the γ -aminobutyric moiety. More generally, the comparison of these results with the activities of the corresponding acids [9] did not prove any superiority of esters.

In contrast, esterification always enhances anthelmintic activity (mainly filaricid). The EC₅₀ values were < 50 μ g/ml for 14 compounds against *M* dessetae at 7 d, whereas the acid analogues of **9a** and **9g** and the two most active compounds **9p** and **9s** were completely ineffective [2]. Examination of the results did not prove any superiority of β -alanine over the γ -aminobutyric moiety.

Table II. NMR data for 9a-w (CDCl₃ or CDCl₃/CD₃ODa at 60 MHz).

Ar—NH—
$$C$$
—NH— (CH_2) $\frac{1}{n}$ COOR

Compound	Aromatic protons	ArN <u>H</u> (s, 1H), N <u>H</u> CH ₂ (t, 1H)	NHC <u>H</u> ₂ (m, 2H), C <u>H</u> ₂ CO (t, 2H), CH ₂ C <u>H</u> ₂ CH ₂ (m, 2H)	Ester R ^b	Others
9a	7.40 (m, 5H)	8.66, 6.90	3.97, 2.83	3.67 (s, 3H)	_
9b	7.33 (m, 5H)	7.92, 6.83	3.87, 2.66	4.15 (q, 2H), 1.18 (t, 3H)	_
9c	7.30 (m, 5H)	8.87, 6.90	3.93, 2.63	4.91 (h, 1H), 1.17 (d, 6H)	_
9d	7.19 (d, 2H), 6.92 (d, 2H)	8.17, 6.63	3.96, 2.68	3.65 (s, 3H)	3.83 (s, 3H, CH ₃ O)
9e	7.37 (d, 2H), 6.93 (d, 2H)	7.90, 6.88	3.92, 2.73	4.16 (q, 2H), 1.37 (t, 3H)	3.83 (s, 3H, CH ₃ O)
9f	7.25 (d, 2H), 6.88 (d, 2H)	8.07, 6.74	3.95, 2.70	4.89 (h, 1H), 1.20 (d, 6H)	3.82 (s, 3H, CH ₃ O)
9 g ª	8.15 (d, 2H), 7.75 (d, 2H)	exch, exch	3.93, 2.75	3.39 (s, 3H)	_
9h	6.85–7.01 (m, 3H)	8.42, 6.85	3.86, 2.76	3.93 (s, 3H)	3.85–3.88 (2s, 6H, 2 CH ₃ O)
9i	6.82–6.95 (m, 3H)	8.40, 6.90	3.83, 2.75	4.15 (q, 2H), 1.25 (t, 3H)	3.85 (2s, 6H, 2 CH ₃ O)
9j	6.83-6.97 (m, 3H)	8.55, 6.45	3.96, 2.71	5.08 (h, 1H), 1.23 (d, 6H)	3.84 (2s, 6H, 2 CH ₃ O)
9k	6.85-6.94 (m, 3H)	8.18, 6.75	3.83, 2.63	3.65 (s, 3H)	4.40 (m, 4H, OCH ₂ CH ₂ O)
91	6.90-6.98 (m, 3H)	8.05, 6.48	4.05, 2.43	5.02 (h, 1H), 1.25 (d, 6H)	4.38 (m, 4H, OCH ₂ CH ₂ O)
9m	7.35 (m, 5H)	8.77, 6.67	3.68, 2.66, 1.93	3.63 (s, 3H)	() , 2 2 /
9n	7.34 (m, 5H)	8.90, 6.95	3.95, 2.65, 1.95	4.12 (q, 2H), 1.27 (t, 3H)	_
90	7.32 (m, 5H)	8.86, 6.67	3.63, 2.63, 1.93	4.93 (h, 1H), 1.20 (d, 6H)	_
9p	7.30 (d, 2H), 6.97 (d, 2H)	8.40, 6.20	3.80, 2.37, 1.97	3.67 (s, 3H)	3.87 (s, 3H, CH ₃ O)
9 q	7.27 (d, 2H), 6.98 (d, 2H)	8.18, 6.23	3.78, 2.37, 1.88	4.15 (q, 2H), 1.23 (t, 3H)	3.85 (s, 3H, CH ₃ O)
9r	7.23 (d, 2H), 6.90 (d, 2H)	8.16, 6.18	3.67, 2.32, 1.85	4.97 (h, 1H), 1.22 (d, 6H)	3.83 (s, 3H, CH ₃ O)
9s ^a	8.10 (d, 2H), 7.79 (d, 2H)	exch, exch	3.83, 2.70, 1.81	3.70 (s, 3H)	_
9ta	8.14 (d, 2H), 7.79 (d, 2H)	exch, exch	3.80, 2.70, 1.81	4.79 (h, 1H), 1.17 (s, 6H)	-
9u	6.95 (m, 3H)	8.15, 6.32	3.83, 2.35, 1.98	4.98 (h, 1H), 1.18 (d, 6H)	3.95 (2s, 6H, 2 CH ₃ O)
9v	6.83 (m, 3H)	8.12, 6.23	3.73, 2.40, 1.96	3.73 (s, 3H)	4.33 (m, 4H, OC <i>H</i> ₂ C <i>H</i> ₂ O)
9w	6.80 (m, 3H)	8.05, 6.42	3.80, 2.33, 1.98	5.00 (h, 1H), 1.26 (d, 6H)	4.38 (m, 4H, OC H_2 C H_2 O)

 $^{{}^{}a}$ In CDCl₃/CD₃OD; b R = CH₃, C_{2} H₅ or i- C_{3} H₇, s = singlet; d = doublet; t = triplet; q = quartet; h = septet.

Table III. Antifungal activity of 9a-w against 36 strains of Candida (MIC in µg/ml).

Compound	C albicans	C krusei	C glabrata	C tropicalis	C parapsilosis
9b YNBG CAS	58 33	48 32	64 32	32 17	32 19
9c YNBG CAS	22 19	16 16	16 16	16 16	16 11
9d YNBG CAS	6 5	6 4	16 16	16 16	12 8
9h YNBG CAS	68 30	32 32	32 16	33 30	37 35
9i YNBG CAS	101 70	128 128	64 64	106 110	68 60
9j YNBG CAS	112 98	64 64	64 64	36 30	48 36
91 YNBG CAS	58 47	32 32	32 32	36 28	30 30
9r YNBG CAS	24 20	32 16	32 32	28 20	24 24
9s YNBG CAS	10 11	8 8	8 8	14 16	18 16
9t YNBG CAS	12 10	8 8	8 4	12 8	16 10
9w YNBG CAS	38 36	64 32	64 32	24 18	28 16
Econazole	11	16	13	28	20

Results are not reported for 9a, 9c, 9e-g, 9k or 9m-q whose MIC > 128 μ g/ml for all strains.

Experimental protocols

Chemistry

General method for preparation of 9a-w

To 0.010 mol of hydrochloride of the appropriate ester in 20 ml water was added 4 ml triethylamine (0.030 mol), and then dropwise 0.010 mol of the appropriate isothiocyanate in 20 ml acetone. The mixture was stirred for 4 h at 40°C, and then evaporated under reduced pressure. The residue was suspended

in water and extracted with methylene chloride. The organic layer was dried on $MgSO_4$ and concentrated under reduced pressure. The crude residue (a white-yellow oil) was triturated with diethyl ether to give a white powder, which was purified by recrystallization in THF.

Physicochemical data for 9a-w

Melting points were determined on a Kofler bank. Yields are given for the final step of the synthesis. Infrared data (in KBr pellets) were measured with a Perkin-Elmer 983-G spectro-

Table IV. Antifungal activity of 9a-w against six strains of Aspergillus and two strains of S brevicaulis (MIC in µg/ml).

	A.C	A (1)	4	S brevicauli
Compound	A fumigatus	A flavus	A niger	S brevicauii
9b				
YNBG	32	64	64	32 32
CAS	16	64 32	16	32
9c				
YNBG	32	64	64 32	48
CAS	32 24	64 32	32	32
9d				
YNBG	6	8	8	8 4
CAS	6 2	8 4	8 4	4
9h				
YNBG	48	64	64	64
CAS	32	32	64 32	32
9i				
YNBG	64	111	64	64
CAS	64	111	64	64
9j				
YNBG	64	> 128	64	64
CAS	32	64	32	64
91				
YNBG	64	> 128	64	64
CAS	64	> 128	32	64
9r				
YNBG	64	111	48	32
CAS	32	64	64	64
9s				
YNBG	12	16	12	8
CAS	4	8	12 2	4
9t				
YNBG	12	8 8	8 8	16 8
CAS	4	8	8	8
9w				
YNBG	48	48	64	64
CAS	48 32	32	64	32
Econazole	4	4	2	64

Results are not reported for 9a, 9c, 9e-g, 9k or 9m-q whose MIC > 128 μ g/ml for all strains.

meter, v (in cm⁻¹): NH 3300-3400 and 3200-3240; CO 1710-1730. NMR data were collected in CDCl₃ or CDCl₃/CD₃OD 4:1 for nitro derivatives which are insufficiently soluble in CDCl₃, at 60 MHz and with Me₄Si as an internal standard. Elemental analyses were in agreement with the accepted norms and are not reported.

Parasitology

Antifungal activity

All compounds were tested against an array of clinical isolates and one reference strain: 25 C albicans strains and one C albi-

cans AFNOR (ATCC 2094), reference one C glabrata, one C krusei, four C parapsilosis, four C tropicalis, two A fumigatus, two A niger, two A flavus and two S brevicaulis.

MIC were performed on two media, YNBG and CAS, with solutions of 9a—w in DMSO/water (10:90) using the method of dilution on gelose [15, 16]. On the same medium, no antifungal activity was noted for the DMSO/water mixture.

Compounds 9a-w were screened in vitro against infectious larvae of an intestinal parasite of rats, N brasiliensis, and infectious larvae of a filaria M dessetae. These two tests were chosen because they tend to detect in vitro activities which are generally confirmed in vivo; experimental procedures have been published previously [12-14].

Table V. In vitro anthelmintic activity of 9a-w derivatives (EC₅₀ in μg/ml).

Compound	L3 of N b	orasiliensis – – – – – – – – – – – – – – – – – –	L3 of M dessetae		
	24 h	96 h	24 h	168 h	
9a	I	I	I	20	
9b	200	120	45	28	
9c	200	95	48	30	
9f	180	80	I	10	
9g	1	I	80	30	
9i	I	110	120	50	
91	I	1	I	35	
90	I	100	I	5	
9p	I	I	120	3	
9q	I	1	32	25	
9r	150	75	50	25	
9s	200	100	80	5	
9u	180	125	100	50	
9w	200	200	125	30	
Tetramisole	1.7	0.2	65	3	

I: inactive compound (EC₅₀ > 200 μ g/ml). Results are not reported for **9d-e**, **9h**, **9j-k**, **9m**, **9t** or **9v** whose EC₅₀ > 200 μ g/ml for all strains. L3 = third larval instar.

References

- Brewer MD, Dorgan RJJ, Manger BR, Mamalis P, Webster RAB (1987)
 J Med Chem 30, 1848–1853
- 2 Parish RC (1977) US Patent 4002761; Chem Abstr 86-13987g
- 3 Shridhar DR, Srinivasa R, Tripath HW, Sai GST (1981) Indian J Chem 20-h 471-474
- 4 Shridhar DR, Ghandi SS, Srinivasa R (1982) Indian J Chem 21-b, 971-972
- Walchshofer N, Minjat M, Tinland B, Jaussaud P, Petavy AF, Paris J (1986) Eur J Med Chem 21, 59-64
- 6 Doré JC, Lacroix J, Lacroix R, Viel C (1987) Eur J Med Chem 22, 109-117
- Wolff ME (1979) Burger's Medicinal Chemistry 4th ed, Wiley Interscience, New York, vol 2, 531-541

- 8 Caujolle R, Amarouch H, Payard M et al (1989) Eur J Med Chem 24, 287-292
- 9 Caujolle R, Payard M, Loiseau PR et al (1991) Eur J Med Chem 26, 723-727
- 10 Bennet JC, Williams JF, Dave V (1988) Parasitol Today 4, 226-228
- 11 Hodgkins JE, Reeves WP (1964) J Org Chem 22, 439-443
- 12 Brienne MJ, Varech M, Leclercq M et al (1987) J Med Chem 30, 2222-2232
- 13 Gayral P, Gueyouche C, Bories C et al (1989) Drug Res 39, 226-230
- Bories C, Loiseau P, Legrand J, Gayral P (1987) Bull Soc Fr Parasitol 5, 75-78
- 15 Mailie M (1985) Doctoral thesis, Université Montpellier I, France
- 16 Cazaux M, Linas MD, Bessieres MH, Recco P, Seguela JP (1980) Bull Soc Mycol Med 9, 259–262